

NMR STUDIES OF 1-PHENYLAZO-3-SUBSTITUTED-2-NAPHTHOLS IN SOLUTION AND IN THE SOLID STATE

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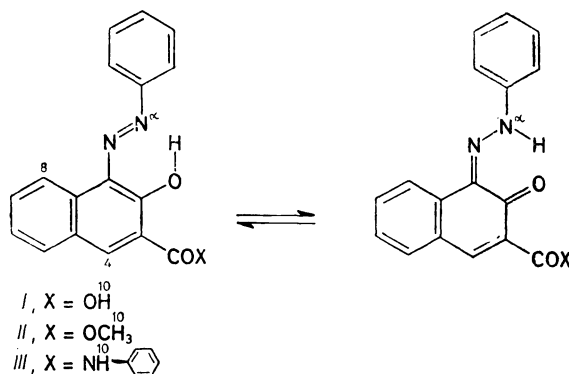
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Dedicated to Professor Otto Exner on the occasion of his 65th birthday.

The ¹H and ¹⁵N NMR spectra of ¹⁵N_α-enriched 1-phenylazo-3-X-2-naphthols, where X = COOH (*I*), X = COOCH₃ (*II*), and X = CONHC₆H₅ (*III*), have been measured in various solvents. The values of ¹J(¹⁵N_α, H) and δ(¹⁵N) indicate that in CDCl₃, C₆D₆, CCl₄, and CD₃NO₂ solutions the compounds *I* and *III* exist practically completely in their hydrazone forms. The hydrazone form is stabilized by the hydrogen bond of COOH or CONH protons to the C(2)=O group. The compound *II* represents an equilibrium mixture of azo and hydrazone forms, since it cannot form a similar hydrogen bond. Moreover, the ¹⁵N NMR spectra of compounds *I–III* have been measured in solid state by the CP/MAS technique. The results indicate the existence of two conformers differing by the conformation of COOCH₃ group in compound *II*, which is supported by the ¹³C CP/MAS NMR spectrum of compound *II*.

The 1-phenylazo derivatives derived from 3-substituted-2-naphthols, particularly those having CONHC₆H₅ or substituted phenylaminocarbonyl group at the 3-position, are important organic pigments with broad technical applications¹. As to real



structure of these compounds in solution² and in solid state³ their hydrazone form is usually preferred or is stated to coexist in a mixture with the azo form both in solution^{4,5} and in solid state⁶.

The aim of the present communication is to apply the NMR spectroscopy in solution and in the solid state to evaluate the azo-hydrazone tautomerism of the ¹⁵N_α-enriched 1-phenylazo-3-substituted 2-naphthols *I–III*. The 1-phenylazo derivatives were selected, because they form solutions still applicable to the measurements of the ¹H and ¹⁵N NMR spectra of ¹⁵N_α-enriched compounds by long-term accumulation. If there are (particularly electron-acceptor) substituents in the phenyl groups, the compounds are quite insoluble in current solvents.

EXPERIMENTAL

The ¹⁵N-enriched compounds *I* and *III* were obtained by azo coupling of the benzenediazonium chloride prepared from ¹⁵N-aniline (96% ¹⁵N) with 3-hydroxy-2-naphthoic acid or 2-hydroxy-naphthalene-3-carboxanilide⁷. Compound *II* was prepared by reaction of acid *I* chloride with methanol⁸.

The ¹H NMR spectra were measured in the standard way at 99.602 MHz with a JNM-FX 100 (JEOL) apparatus in 10 mm NMR tubes. Due to low solubility, the samples for the measurement were saturated solutions in CDCl₃, C₆D₆, CCl₄, and CD₃NO₂. The temperature of the heating or cooling gas was measured with a thermocouple. The ¹H chemical shifts were related to internal tetramethylsilane ($\delta = 0.00$). The ¹H NMR spectra of the ten times diluted samples were measured at 400.13 MHz with a Bruker AM 400 apparatus. The ¹⁵N NMR spectra of compounds *I–III* were obtained at 10.095 MHz with the JNM-FX 100 apparatus using the same samples as for the measurements of ¹H NMR spectra. The measurement conditions: spectral width 5 000 Hz, 8 K memory, pulse repetition 4 s, proton noise decoupling. The ¹⁵N chemical shifts are related to external nitromethane (25% ¹⁵N, $\delta = 0.0$).

The solid-state ¹⁵N NMR spectra were measured at 20.28 MHz with a Bruker MSL 200 spectrometer using the CP/MAS technique^{9,10} at room temperature. Alternation of the spin temperature in the rotating frame to suppress artifacts¹¹ and the flip-back pulse¹² were used. The samples were packed in double-bearing Al₂O₃ rotors supplied by Bruker. The magic angle was adjusted by minimization of the line width of the carbonyl carbon of glycine. Typical measurement conditions were as follows: 90° r.f. pulse: 5.5 μ s, contact time 6 ms, pulse repetition time 5 s, number of transients 320, MAS frequency 5 kHz, spectral width 20 kHz, acquisition time 51 ms. The ¹⁵N chemical shifts were referenced to solid NH₄Cl by sample replacement and recalculated to nitromethane using $\delta(\text{NH}_4\text{Cl}) = -352.5$ (ref.¹³). The solid state ¹³C CP/MAS NMR spectrum was taken on the same instrument at 50.32 MHz. The measurement conditions were as follows: 90° r.f. pulse: 5.5 μ s, contact time 1 ms, pulse repetition time 5 s, number of transients 2 400, MAS frequency 4.5 kHz, spectral width 20 kHz, acquisition time 26 ms. The ¹³C chemical shifts were referred to the carbonyl carbon atom of glycine ($\delta = 176.03$) by sample replacement. In all the cases positive values of chemical shifts denote downfield shifts.

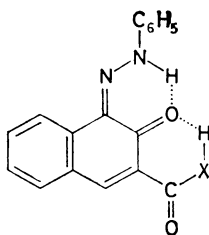
RESULTS AND DISCUSSION

NMR spectroscopy represents a very appropriate method for investigation of azo-hydrazone tautomerism^{14,15}. Several NMR characteristics have been proposed

for its evaluation¹⁵. The coupling constants $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{exp}}$ and chemical shifts $\delta(^{15}\text{N})$ of the $^{15}\text{N}_\alpha$ -enriched substances are applicable to solution of our problem of the measurements in solution because of the extremely low solubility of the substances measured. Table I presents the values of $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{exp}}$, the ^1H and $^{15}\text{N}_\alpha$ chemical shifts of compounds *I–III* in various solvents and at various temperatures as well as the calculated contents of hydrazone forms in compounds *II* and *III*. The proton of N_αH group is split by the ^{15}N isotope (the spin number $I = 1/2$) into a doublet, the magnitude of the $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{exp}}$ coupling constant enabling the determination of azo and/or hydrazone forms content in the azo dyestuffs measured. The coupling constant $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{exp}}$ of compound *I* is independent of temperature and of the solvent used and corresponds to the compound existing in its hydrazone form. On the other hand, the magnitude of the coupling constants $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{exp}}$ of compound *II* depends on both the temperature and the solvent used, the coupling constants being distinctly smaller than those of compound *I*. Equation (1) was used to estimate the contents of the hydrazone form^{16,17}:

$$\% \text{ hydrazone form} = 100 ({}^1J(^{15}\text{N}_\alpha, \text{H})_{\text{exp}} / {}^1J(^{15}\text{N}_\alpha, \text{H})_{\text{H}}), \quad (1)$$

where $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{exp}}$ is the value measured for the respective tautomeric system and $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{H}}$ is that corresponding to the 100% content of hydrazone form. For the compounds without the second hydrogen bond (vide infra) it was suggested¹⁷ to adopt the value $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{H}} = 96$ Hz. The content of hydrazone form in compound *II* varies from 60 to 78%, depending on the temperature and the solvent used. The coupling constants $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{exp}}$ in compound *III* are very similar to those in compound *I*, but they do show a certain dependence on temperature and solvent. Equation (1) was adopted also for the estimation of the hydrazone form content in compound *III*, but the coupling constant taken was $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{H}} = 92.4$ Hz (see compound *I*), since the compounds able to form the second hydrogen bond (vide infra) have lower $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{H}}$ values¹⁸ as compared with those unable to form this second hydrogen bond. The main difference between compounds *I*, *III* and the



IV, X = O or NC_6H_5

TABLE I

Values of $^1J(^{15}\text{N}_g, \text{H})_{\text{exp}}$, ^1H , and ^{15}N chemical shifts of compounds I—III in various solvents at various temperatures and calculated content of hydrazone form in compounds II and III

Compound	Solvent	Temperature, K	$^1J(^{15}\text{N}_g, \text{H})_{\text{exp}}$ Hz	% ^b	$\delta(^{15}\text{N}_g)$	% ^d	$\delta(\text{NNH})$	$\delta(\text{H}(4))^e$	$\delta(\text{H}(8))^e$	$\delta(\text{H}(10))$
I	CDCl_3	300	92.5	c	-173.8	c	16.56	8.88	8.45	13.70
I	CDCl_3	330	92.3	c	—	—	16.54	8.89	8.47	13.50
I	C_6D_6	300	92.5	c	-178.0	c	16.16	8.96	8.33	13.64
I	CD_3NO_2	300	92.3	c	—	—	16.33	8.76	8.57	13.45
II	CDCl_3	300	74.8	77.9	-140.5	76.5	16.89	8.41	8.46	3.96
II	CDCl_3	330	71.0	74.0	-131.4	73.1	16.74	8.33	8.41	3.86
II	C_6D_6	300	61.5	64.1	-112.5	66.3	16.85	8.28	8.56	3.76
II	C_6D_6	350	58.1	60.5	-101.9	62.3	16.50	8.27	8.56	3.79
II	CCl_4	300	58.2	60.6	—	—	16.57	8.28	8.50	3.91
III	CDCl_3	300	90.8	98.3	-178.1	c	16.81	8.96	8.37	11.57
III	CDCl_3	330	89.8	97.2	-175.9	c	16.79	8.96	8.43	11.46
III	C_6D_6	300	90.8	98.3	-180.5	c	16.46	9.08	8.25	11.57
III	C_6D_6	350	88.4	95.7	-176.2	c	16.50	9.06	8.28	11.45
III	CCl_4	300	90.5	97.9	—	—	16.86	8.94	8.43	11.25

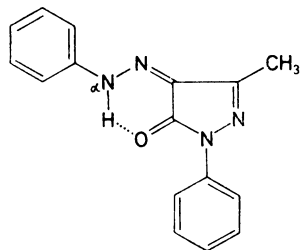
^a (± 0.3 Hz); ^b the percentage of hydrazone form calculated according to Eq. (1); ^c see the text; ^d the percentage of hydrazone form calculated according to ref.¹⁹; ^e the chemical shifts ^1H of the other aromatic protons are found within the interval of 7.10—8.00 ppm.

compound *II* consists in the fact that compounds *I* and *III* form the second intramolecular^{5,15} hydrogen bond (*IV*) which is very strong ($\delta(^1\text{H})_{\text{COOH}}$ of compound *I* is 13.45 to 13.70), whereas compound *II* does not. This second intramolecular hydrogen bond represents a very significant stabilizing factor of the hydrazone form. The content of hydrazone form is practically 100% in compounds *I* and *III*, whereas that in compound *II* is roughly the same as in 1-phenylazo-2-naphthol (which exists as 70% 1,2-naphthoquinone 1-phenylhydrazone in deuteriochloroform¹⁹).

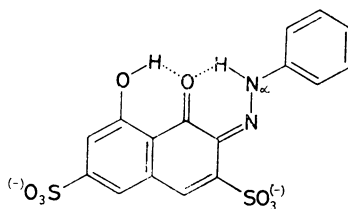
The ¹H chemical shifts of COOH and CONH protons in compounds *I* and *III* remain unaffected even after a tenfold dilution of the saturated solutions of the substances in deuteriochloroform, which was proved by measurements of ¹H NMR spectra at 400.13 MHz, and this fact can be considered – with respect to the very low concentrations used – to prove the intramolecular character of these hydrogen bonds.

At 99.602 MHz it is possible to assign – out of the aromatic protons – the ¹H chemical shifts of the H(4) protons (giving a singlet) and the H(8) protons at the peri position with regard to phenylhydrazone group. The other signals of aromatic protons form strongly overlapping multiplets within the region from 7.10 to 8.00 pm.

The ¹⁵N chemical shifts in compounds *I* and *III* have the value of –173.8 or are shifted even more upfield. The values are typical of almost 100% hydrazone compounds with two intramolecular hydrogen bonds. The value $\delta(^{15}\text{N}_\alpha) \approx -205$ of compound *V* is used as a standard value for a 100% hydrazone form with one intramolecular hydrogen bond, whereas the value $\delta(^{15}\text{N}_\alpha) = -186.3$ was measured¹⁸ for the compound *VI* which also exists as 100% hydrazone form, however, stabilized



V



VI

by the second intramolecular hydrogen bond. The hydrazone form content in compound *II* was calculated with the use of $\delta(^{15}\text{N}_\alpha)$ values by the procedure given elsewhere¹⁹. Table I shows a reasonable agreement between the values of hydrazone form content determined in compound *II* by means of $^1J(^{15}\text{N}_\alpha, \text{H})_{\text{exp}}$ and $\delta(^{15}\text{N}_\alpha)$.

For the compounds of the type *I–III*, which – if containing electron-acceptor substituents – are insoluble in current solvents, it is important that we might know the content of the hydrazone forms in solid phase. Earlier we described²⁰ the ap-

plication of ^{15}N CP/MAS NMR spectra to estimation of the hydrazone form content in solid state of model azo dyestuffs. The same technique has now been used to measure the solid compounds *I–III* at room temperature. The ^{15}N CP/MAS NMR spectra of the $^{15}\text{N}_\alpha$ -enriched compounds *I–III* are shown in Fig. 1. The compounds *I* and *III* exhibit one signal with the chemical shifts of -190.8 and -199.9 , respectively. As compared with the values obtained in solution, the signals obtained by the measurement in solid state are shifted upfield by c. $10–17$ ppm. Similar differences were also found with the data of ref.²⁰ and were assigned to the differences in $\delta(^{15}\text{N})$ of both the standard and the compounds in solution and solid state. Thus in solid state no distinct change was observed in the content of hydrazone form in compounds *I* and *III*, and these compounds exist also in solid state as 100% hydrazone forms. This result agrees with the X-ray diffraction data (see ref.²¹ and the literature quoted therein) of similar substances which were proved to exist as planar structures in hydrazone forms with intramolecular hydrogen bonds. From the energy dispersive X-ray spectra⁶ of compound *III* it was concluded that the solid substance exists as 52% hydrazone form and 48% azo form. This conclusion contradicts our results obtained from the ^{15}N CP/MAS NMR spectra. The difference of ^{15}N chemical shifts between the azo and hydrazone forms is c. 275 ppm for N_α . If compound *III* had the composition given in ref.⁶, then the signal of N_α would have to lie in the region about -30 ppm. Therefore we suppose that the method

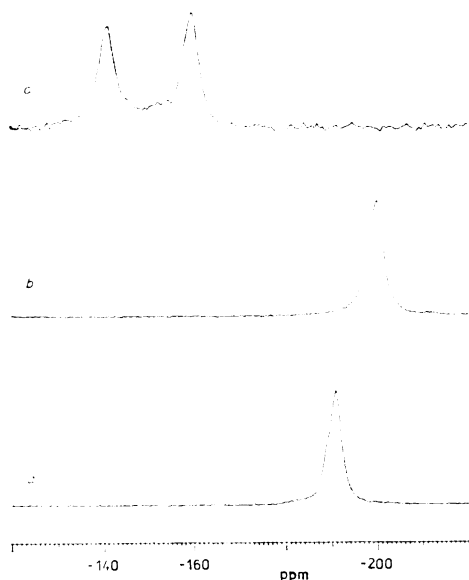


FIG. 1
The ^{15}N CP/MAS NMR spectra of compounds *I* (a), *II* (c), and *III* (b)

adopted⁶ is inappropriate for characterization of the azo-hydrazone tautomerism. The method of energy dispersive X-ray spectra should be used to measure substances known to exist practically either as 100% azo or 100% hydrazone form, whereby the method would be verified.

The ¹⁵N CP/MAS NMR spectrum of compound *II* (Fig. 1c) exhibits two signals with the chemical shifts of -141.2 and -159.8. Two ¹⁵N chemical shifts were also observed in the ¹⁵N CP/MAS NMR spectrum of 1-phenylazo-2-naphthol in solid state²⁰ where the difference between the ¹⁵N chemical shifts varied (depending on the temperature) within the limits of 3.4–5.8 ppm and was ascribed to crystallographical nonequivalence in the unit cell. A difference of 3.4 ppm was described³ for calcium(II) salt of (1-sulfo-4-methylphenylazo)-2-hydroxynaphthalene-3-carboxylic acid. In the case of compound *II*, however, the difference is markedly greater (18.6 ppm). The signals of ¹⁵N CP/MAS NMR spectrum lying at -141.2 and -159.8 ppm can be interpreted as signals of two different azo-hydrazone forms in

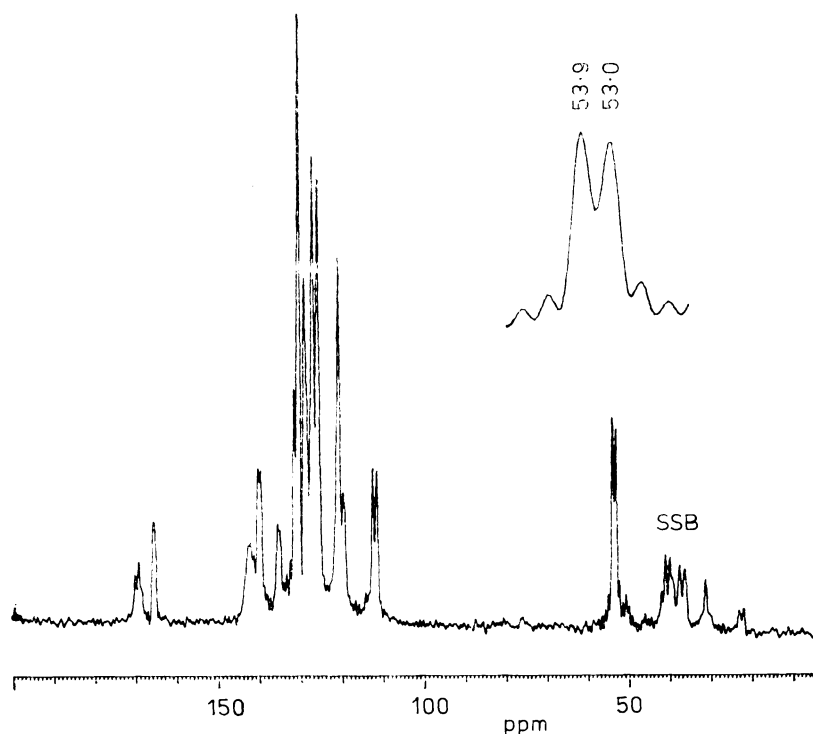
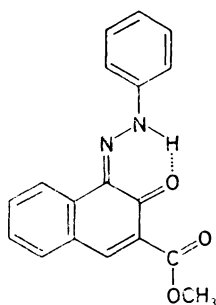
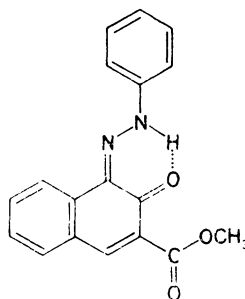


FIG. 2
The ¹³C CP/MAS NMR spectrum of compound *II* (SSB denote spinning side-bands)

compound *II*, the proton exchange must be rapid judging from the signals half-widths. These two tautomerisms could arise either from the fact that compound *II* is composed of an amorphous and a crystalline parts or from a possibility of two conformations of compound *II* stable in solid state. In order to verify the existence of two stable conformers of compound *II*, we measured the ^{13}C CP/MAS NMR spectrum of this compound. The spectrum is given in Fig. 2, and it can be seen that it is composed of two sets of signals. The most striking splitting of signals is that of OCH_3 group. The bands of both the components are approximately of the same intensity, which is analogous to the two bands in the ^{15}N CP/MAS spectrum. All this agrees with the idea that there are two stable conformers present, formed by rotation around the $\text{C}-\text{COOCH}_3$ bond. As, according to the X-ray analysis, molecules of the type *II* are planar, our two conformers obviously are those given in formulas *VII* and *VIII* for the hydrazone form.



VII



VIII

Different situations with respect to interatomic distances ($\text{N}-\text{H}$, $\text{O}-\text{H}$) and electronic densities at the individual atoms then cause changes in the content of the tautomeric forms, the difference of 18.6 ppm for N_α atom corresponding to a 6.8% change in the azo or hydrazone form content (for a difference²⁰ of c. 275 ppm between "100%" azo and hydrazone compounds for N_α atom). The content of hydrazone forms determined from Eq. (1) and ref.²⁰ is c. 70 and 77% for the individual conformers of compound *II*. This content agrees well with the value of the hydrazone content determined in solution of compound *II* (Table I). Univocal evidence in support of the conformers *VII* and *VIII* could be provided by X-ray analysis.

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